Fertilization and carbon sequestration

Vladimír Šimanský

Slovak University of Agriculture in Nitra, Slovakia

The effect of fertilization on the dynamic change of soil organic matter (SOM) in loamy Haplic Luvisol was studied. In 2008– 2011, soil samples were taken from following treatments: 1. C - non-fertilized, 2. PR + NPK - plant residues together with NPK fertilizers, and 3. NPK – NPK fertilizers. The results showed that the content of soil organic carbon in water-stable microaggregates (SOC in WSA_m) increased by 11% and by 13% in PR + NPK and NPK treatments, respectively. The ratios of SOC in WSA_/SOC in bulk soil in the NPK fertilized treatment and in PR + NPK were 14% and 4% higher than in the non-fertilized treatment, respectively. Overall the ratios of SOM in WSA/SOM in bulk soil were higher in macro-aggregates than microaggregates. In fertilized treatments, the statistical significant changes in dynamics of labile carbon in water-stable macroaggregates (C, in WSAma) and in WSAma 0.5-3 mm were observed. In fertilized treatments (PR + NPK and NPK) there were observed significant decrease of the C_{L} in WSAma. The ratios C_{L} in WSA_{mi} also WSA_{ma}/ C_{L} in bulk soil decreased due to ploughed plant residues together with NPK fertilizers, the ratio of C_L in WSA_{ma}/ C_L in soil decreased due to added only NPK fertilizers also.

Keywords: soil organic matter, water-stable aggregates, plant residues, soil management practices

1. Introduction

Increasing problems concerning the environmental quality in arable landscapes, and the long-term productivity of agro-ecosystems, have emphasized a need to develop and improve management strategies that maintain and protect soil function and resources. If a farmer wants to be successful, they must have a good knowledge of the soil and 'understand'it. Using the knowledge of chemistry and soil physics, one is able to design the proper soil management practices, ensuring its protection and sustainable production.

SOM is an important aspect of agricultural soil quality and soil ecology (Balashov and Buchkina, 2011; Gaida et al., 2013). SOM is a dynamic entity. The amount of organic matter in a given soil can increase or decrease depending on numerous factors including climate, vegetation type, nutrient availability, disturbance, land use, and management practices (Six and Jastrow, 2002). A stabilization and protection of SOM is controlled by the following mechanisms: association of SOM with clay minerals and Fe and Al oxides, sequestration into macro and micro pores of soil aggregates; and biochemical stabilization (Six et al., 2002; Chenu and Plante, 2006; von Lützov et al., 2008). Nevertheless, soil has a limited capacity of saturation by SOM within soil mineral matrix and aggregates (Eusterhues et al., 2003). Soil aggregation is not only an important process of carbon sequestration,

but it is a key factor controlling the quality of arable soils, as it plays an important role in the formation of their optimal physical conditions. It can be affected by the whole complex of natural and anthropogenic influences. Of all the anthropogenic influences, fertilization has one of the greatest impacts. Fertilizers, in a broad sense, include all materials that are added to soils to increase the growth, yield, quality, or nutritive value of crops. Fertilizers may affect the soil and plant growth in a number of different ways (Millar et al., 1962).

The aim of this paper was to study how fertilization influences SOM dynamics. We compare the SOM in soil, as well as in water-stable aggregates (WSA), in a Haplic Luvisol subjected to ploughed plant residues, together with NPK fertilizer or NPK treatment during the years 2008-2011.

2. Material and methods

An experiment is situated at the Dolná Malanta (48019'00" N; 18° 09' 00" E) where, in 1994, the Department of Plant Production of SAU Nitra established a long-term field experiment, which is still running. The experimental site location is to the east of Nitra city, on the Žitavská upland. The experimental area is flat, with a slight incline southwards. The geological substratum consisted of little previous rocks with high quantities of fine materials. Young Neogene deposits were composed of various

*Correspondence: Vladimír Šimanský, Slovak University of Agriculture in Nitra, Faculty of Agrobiology and Food Resources, Department of Soil Science, Trieda Andreja Hlinku 2, 949 76 Nitra, Slovak Republic, e-mail: vladimir.simansky@uniag.sk

clays, loams, sand gravels on which loess was deposited in the Pleistocene Epoch. Soil was classified according to World Reference Base for Soil Resources (WRB, 2006) as the loamy haplic Luvisol. Before the experiment the soil contained 360.4 g kg⁻¹ of sand, 488.3 g kg⁻¹ of silt and 151.3 g kg⁻¹ of clay. Soil carbon content was 12.9 g kg⁻¹, while the cation exchange capacity was 147.18 mmol kg⁻¹ and base saturation percentage was 92.6%. On average, the soil active pH was 6.96. The experimental site has a mean annual temperature of 9.8 °C, and a mean annual precipitation of 573 mm.

In 2008–2011, twice a year (spring and autumn), the soil samples were taken from the depth 0–0.2 m from following treatments: C – non-fertilized, PR + NPK – plant residues ploughed to the soil together with NPK fertilizers, and NPK – added NPK fertilizers. The doses of fertilizers were: N 80 kg ha⁻¹, P (P_2O_5) 45 kg ha⁻¹ and K (K_2O) 72 kg ha⁻¹. In 2008 in experimental plot was sown red clover (*Trifolium pratense* L.), in 2009 winter wheat (*Triticum aestivum* L.), in 2010 pea (*Pisum sativum* L. subsp. Hortense (Neitr.) and in 2011 maize (*Zea mays* L.).

Soil samples for the determination of SOM parameters as soil organic carbon content - SOC (Dziadowiec and Gonet, 1999) and labile carbon content – C₁ (Loginow et al., 1987) in bulk soil were collected from each sampled zone (included all treatments of fertilization) and mixed to an average sample. Samples from the same sampled zones for the determination of SOM parameters in individual size fraction of WSA (SOC and C, – according to above mentioned methods) were taken with the aid. Large clods were gently broken up along natural fracture lines, and samples were then air-dried at the laboratory temperature. Before the determination of WSA, all soil samples were sieved to provide a range of aggregate sizes (>7, 7-5, 5-3, 3-1, 1-0.5, 0.5-0.25, <0.25 mm). These size classes of air-dried aggregates were used for the determination of size classes of WSA by the Baksheev method (Vadjunina and Korchagina, 1986), while aggregates in size fractions more than 0.25 mm are macro-aggregates (WSA $_{ma}$) and less than 0.25 mm are micro-aggregates (WSA_{mi}). The share of macroaggregates in size from 0.5 to 3 mm is important from the agronomical point of view.

The statistical treatment of the data was performed with the use of the Statgraphics Centurion XV.I (Statpoint Technologies, Inc., USA). Treatment differences (ANOVA) were considered significant at *P*-values <0.05 by the LSD multiple-range test. Non-linear regression analyses were performed to quantify the dynamic change of SOM in bulk soil as well as in WSA.

3. Result and discussion

Results from 2008 to 2011 were evaluated (Table 1). Overall, application of fertilizers did not significantly influence the SOC because a significant part of the SOC is formed from the stable fraction of organic matter and it has been turned over thousands of times over the years (Haynes, 2005; Richter et al., 2007; Šimanský et al., 2013). As presented in the results (Table 1), there was no significant change in the content of C₁ due to fertilization. This is surprising because several previous studies demonstrate that labile C pools recover in shorter time frames than total SOC (Carpenter-Boggs et al., 2003; Dou et al., 2008; Šimanský, 2013). In the PR + NPK treatment, plant residues added to the soil together with NPK fertilizers increased (no significant, P > 0.05) the SOC in soil, but the content of C, was the same in comparison to the control (nonfertilized). On the other hand, in NPK by 4% compared with the control, there was decreased content of C, in the soil. SOM can be sequestrate into macro and micro pores of soil aggregates (Six et al., 2000; von Lützov et al., 2008) therefore the next important objective of this study was to test the effect of fertilization on re-distribution of SOM in WSA. There were substantial differences ($P \le 0.05$) in SOC in the WSA (mainly in WSA_{mi}) between the fertilized treatments and the non-fertilized treatment. Tisdall and Oades (1980) and Kurakov and Kharin (2012) found greater concentrations of organic C in macroaggregates than in micro-aggregates and suggested that it is due to decomposing roots and hyphae within macro-aggregates. Elliott (1986) suggested that macroaggregates have elevated C concentrations because of the organic matter binding micro-aggregates into macroaggregates and that this organic matter is "qualitatively more labile and less highly processed" than the organics stabilizing micro-aggregates. For the investigated period, the content of SOC in WSA_{mi} increased by 11% and 13% due to ploughed plant residues together with NPK fertilizers and only NPK fertilizers, respectively. In opposition to this, the results of Šimanský (2013) in Rendzic Leptosol showed a decrease of SOC in WSA_{mi} due to the application of higher doses of NPK (120 N kg ha⁻¹, 55 P kg ha⁻¹ and 195 K kg ha⁻¹). In PR + NPK treatment, higher content of SOC by 5% and by 4% was observed in WSA_{ma} and WSA_{ma} size fractions from 0.5-3 mm, respectively. On the other hand in NPK, this trend was the opposite. Fertilizer use improves residue quantity and quality, but this does not necessarily increase SOC pool. However, fertilizers may also decrease SOC concentration in compared to unfertilized soil (Halvorson et al., 2002). Increased ionic concentration in fertilized soil can be the reason for the increase in susceptibility to clay dispersion. This also has a direct impact on the stability of aggregates decrease (Whalen and Chang, 2002), with subsequent decrease in SOC in aggregates because of its low physical protection. Overall, the C₁ content decreased in WSA due to the application of NPK fertilizers. As presented by Neff et al. (2002), the availability of nitrogen from fertilizers

Table 1	Contents o	f soil orgà	anic and la	bile carbo.	n in bulk soil,	in water	r-stable	aggrega	ates and th	neir ratic	suc					
Treatment	SOC	ِ َں		SOC in WS.	'A in g kg ^{.1}		ر in	WSA in	mg kg¹		SOC in W	SA/SOC ii	n bulk soil	C _L in V	VSA/C _L in	bulk soil
	in g kg ⁻	in mg kg	WSA	ni WSA	^{na} WSA _{ma} 0.	5-3 W	'SA _{mi} 1	WSA _{ma}	WSA _{ma} 0.	5-3 W	SA _{mi} V	VSA _{ma}	WSA _{ma} 0.5-3	WSA _{mi}	WSA _{ma}	WSA _{ma} 0.5–3
υ	12.5a	1783a	9.90 ₈	12.85	a 12.8a	16	616a	1862a	1836a	Ö	.80a	1.03a	1.02a	0.91a	1.04a	1.03a
PR + NPK	13.3a	1783a	11.0k) 13.5k	b 13.3a	16	616a	1875a	1843a	Ö	83a	1.03a	1.01a	0.91a	1.05a	1.03a
NPK	12.4a	1719a	11.2b) 12.4 _č	a 12.1a	10	584a	1773a	1722a	Ö	.91a	1.00a	0.98a	0.92a	1.03a	1.00a
	SOC – soil orc aggregates, V	ganic carbc VSA _{ma} – wa	on, C _L – labil ter-stable m	e carbon, SO lacro-aggreg	OC in WSA – soil Jates, WSA _{ma} 0.5	organic ca i–3 – watei	arbon in w r-stable m	vater-stab nacro-agg	ole aggregat Jregates in s	tes, C_ in \ size fraction	WSA – labil on from 0.5	e carbon ir to 3 mm, 0	ı water-stable ago C – control, PR + I	Jregates, W VPK – plan	/SA _{mi} – wat t residues	er-stable micro- oloughed to the
	soil together Different lett	with NPK f. ers betwee	ertilizers, NF n columns (,	ծK – added N a, b) indicate	IPK fertilizers, e that treatment	: means are	e significa	antly diffe	rent at <i>p</i> ≤0).05 accor	ding to LSI) multiple-	range test			
Table 2	Dynamics c	of soil org	anic and l	abile carbc	on in bulk soil	l in indivi	idual tre	eatment	s of fertiliz	ration						
Treatment	Crop	Š	Red clove	er (2008)	Winter whe	at (2009	-	⁹ ea (201	(0	Maize (2011)	Pol	ynomial mode		R ² F	robability
_	time of sar	mpling	spring	autumn	spring	autumn	n sprii	ng au	tumn sp	oring	autumn					
								soc							-	
υ			12.6	12.6	12.3	12.2	11.	6	12.9	12.0	13.6	y = 0.($17x^2 - 0.57x + 13$	3.3 0.	.481	n.s.
PR + NPK			11.5	14.3	12.6	14.5	11.	6	12.0	16.0	13.2	y = -0.	$02x^2 + 0.33x + 1$	2.2 0.	.092	n.s.
NPK			11.6	11.9	11.8	13.0	11.	7	12.6	14.1	12.1	y = -0.	$03x^2 + 0.44x + 1$	1.1 0.	.311	n.s.
								ບ່								
υ			1474	2205	1800	1848	198	30 2	002 1	1232	1724	y = -28.	1 <i>x</i> ² + 224.7 <i>x</i> + 1	489 0.	.248	n.s.
PR + NPK			1395	1530	1800	2138	203	36 2	700 1	1317	1351	y=-73	$2x^{2} + 673.6x + 0$	519 0.	.551	0.05
NPK			1418	1485	1710	1822	201	9 2	216 1	523	1556	y= -42	$.6x^2 + 417.6x + $	926 0.	.634	0.05

SOC - soil organic carbon, C₁ - labile carbon, C - control, PR + NPK - plant residues ploughed to the soil together with NPK fertilizers, NPK - added NPK fertilizers, n.s. - non-significant.

	Crops	Red clo	ver (2008)	Winter w	heat (2009)	Pea (2010)	Maize	(2011)	Polynomial model	R ²	Probability
	time of sampling	spring	autumn	spring	autumn	spring	autumn	spring	autumn			
					U)	OC in WS	\mathbf{A}_{mi}					
υ		7.90	14.5	8.70	10.7	8.60	9.50	8.90	10.7	$y = 0.02x^2 - 0.23x + 10.6$	0.014	n.s.
PR + NPK		6.30	11.0	13.3	17.6	8.90	11.5	8.80	10.7	$y = -0.41x^2 + 3.7x + 4.6$	0.345	n.s.
NPK		15.8	7.80	11.2	14.1	10.4	10.2	9.80	10.0	$y = 0.07x^2 - 1.06x + 14.2$	0.197	n.s.
					сı	OC in WS	A_{ma}					
υ		10.7	14.0	13.5	15.6	12.4	12.4	11.1	13.0	$y = -0.16x^2 + 1.37x + 10.7$	0.252	n.s.
PR + NPK		11.5	15.3	14.1	17.6	11.9	13.4	12.1	12.4	$y = -0.21x^2 + 1.68x + 11.3$	0.307	n.s.
NPK		9.90	11.0	13.8	14.5	11.0	11.9	14.5	12.5	$y = -0.13x^2 + 1.51x + 8.9$	0.337	n.s.
					soc	in WSA _{ma}	0.5–3					
υ		9.70	13.8	13.6	16.4	12.2	12.3	11.4	12.6	$y = -0.24x^2 + 2.11x + 9.2$	0.345	n.s.
PR + NPK		10.4	14.4	14.2	18.5	10.9	13.4	12.1	12.3	$y = -0.26x^2 + 2.28x + 9.8$	0.266	n.s.
NPK		10.6	10.7	13.3	12.8	10.5	12.2	14.7	12.3	$y = -0.04x^2 + 0.70x + 10.1$	0.286	n.s.
						C _L in WSA	m					
υ		1710	1620	1642	1631	1508	1688	1665	1464	$y = -1.08x^2 - 7.89x + 1679$	0.253	n.s.
PR + NPK		1800	1440	1699	2385	1575	1654	1175	1199	$y = -37.21x^2 + 257.8x + 1405$	0.466	n.s.
NPK		1485	1890	1215	1980	1800	1575	1350	1373	<i>y</i> = -23.95 <i>x</i> ² + 184.8 <i>x</i> + 1363	0.251	n.s.
						C _L in WSA	ma					
υ		1782	1841	1545	1800	1928	1967	1549	1772	$y = -5.41x^2 + 47.02x + 1699$	0.030	n.s.
PR + NPK		1892	1947	1961	2453	1895	1878	1501	1469	$y = -37.39x^2 + 265.13x + 1635$	0.689	0.05
NPK		1920	1844	2070	1941	1826	2089	1565	1642	$y = -17.67x^2 + 118.59x + 1779$	0.500	0.05
					์ บ	in WSA _{ma} (0.5–3					
υ		1917	1755	2025	1935	1777	1929	1592	1761	$y = -7.91x^2 + 43.19x + 1844$	0.320	n.s.
PR + NPK		1845	1875	1823	2400	1774	1920	1662	1443	$y = -33.01x^2 + 246.94x + 1573$	0.561	0.05
NPK		1549	1860	1463	1755	1770	2040	1522	1816	$y = -7.13x^2 + 87.04x + 1512$	0.114	n.s.

Ireatment time of samplir C C PR + NPK PR + NPK C PR + NPK PR + NP	ng spring 0.63 0.63 1.36 0.55 0.63 0.85 1.36 1.36 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85	autumn 1.15 0.77 0.66	spring	autumn	spring				-		
C PR+NPK NPK C C C C PR+NPK NPK NPK PR+NPK	0.63 0.55 1.36 1.36 0.85 0.85 0.85 0.85 0.85 0.85 0.85	0.77			· · · · · · · · ·	autumn	spring	autumn			
PR+NPK NPK C C C PR+NPK NPK NPK NPK NPK PR+NPK	0.55 1.36 0.85 1.00 0.85 0.85 0.85 0.85 0.85	0.77 0.66	0.71	0.88	0.72	0.74	0.74	0.79	$y = -0.003x^2 + 0.02x + 0.80$	0.042	n.s.
NPK C PR+NPK NPK NPK C C C C	1.36 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85	0.66	1.06	1.21	0.75	0.96	0.55	0.81	$y = -0.03x^2 + 0.27x + 0.39$	0.393	n.s.
C PR+NPK NPK NPK C C PR+NPK	0.85 1.00 0.85 0.85 0.85 0.85 0.85		0.95	1.08	0.89	0.81	0.70	0.83	$y = 0.10x^2 - 0.13x + 1.27$	0.324	n.s.
C PR+NPK NPK NPK C C C C	0.85 1.00 0.85 0.85 0.85 0.85			SOC in WS.	A _{ma} /SOC	in bulk soil					
PR+NPK NPK NPK C C PR+NPK	1.00 0.85 0.85 0.85 0.85	1.11	1.10	1.28	1.04	0.96	0.93	0.96	$y = -0.02x^2 + 0.16x + 0.80$	0.474	n.s.
NPK NPK C PR+NPK	0.85 0.85 0.77 0.77 0.90	1.07	1.12	1.21	1.00	1.12	0.76	0.94	$y = -0.01x^2 + 0.10x + 0.93$	0.462	n.s.
NPK C PR+NPK	0.85	0.92	1.17	1.12	0.94	0.94	1.03	1.03	$y = -0.01x^2 + 0.09x + 0.81$	0.234	n.s.
C PR+NPK	0.90	0.92	1.17	1.12	0.94	0.94	1.03	1.03	$y = -0.01x^2 + 0.09x + 0.81$	0.234	n.s.
C PR+NPK	0.90			SOC in WSA _{ma}	0.5-3/S(OC in bulk	soil				
PR + NPK	0.90	1.10	1.11	1.34	1.03	0.95	0.95	0.93	$y = -0.02x^2 + 0.21x + 0.68$	0.507	0.05
		1.01	1.13	1.28	0.92	1.12	0.76	0.93	$y = -0.02x^2 + 0.15x + 0.80$	0.382	n.s
NPK	0.91	06.0	1.13	0.98	06.0	0.97	1.04	1.02	$y = -0.002x^2 + 0.02x + 0.91$	0.117	n.s
				C _L in WS	A _m /C _L in	bulk soil					
U	1.16	0.73	0.91	0.88	0.76	0.84	1.35	0.85	$y = 0.02x^2 - 0.14x + 1.14$	0.144	n.s
PR + NPK	1.29	0.94	0.94	1.12	0.77	0.61	0.89	0.89	$y = 0.02x^2 - 0.21x + 1.43$	0.565	0.05
NPK	1.05	1.27	0.71	1.09	0.89	0.71	0.89	0.88	$y = 0.01x^2 - 0.12x + 1.25$	0.303	n.s
				C _L in WS.	A _{ma} /C _L in	bulk soil					
U	1.30	0.84	1.15	1.05	0.92	1.04	1.27	0.95	$y = 0.01 x^2 - 0.09 x + 1.23$	0.083	n.s
PR + NPK	1.36	1.27	1.09	1.15	0.93	0.70	1.14	1.09	$y = 0.02x^2 - 0.25x + 1.64$	0,.614	0.05
NPK	1.26	1.24	0.90	0.99	0.95	0.89	1.02	1.14	$y = 0.02x^2 - 0.24x + 1.51$	0.787	0.01
				C _L in WSA _m	0.5-3/C	in bulk soi	_				
υ	1.30	0.80	1.13	1.05	0.90	0.96	1.29	1.02	$y = 0.01x^2 - 0.13x + 1.27$	0.145	n.s
PR + NPK	1.32	1.23	1.01	1.12	0.87	0.71	1.26	1.07	$y = 0.03x^2 - 0.25x + 1.59$	0.486	n.s
NPK	1.09	1.25	0.86	0.96	0.88	0.92	1.00	1.17	$y = 0.02x^2 - 0.20x + 1.36$	0.547	0.05

© Slovak University of Agriculture in Nitra

can increase the rate of decomposition of labile organic substances. The ratio of SOC in WSA/SOC in bulk soil was evaluated, and the ability of carbon sequestration to exist inside of soil aggregates was recorded (Table 1). In the non-fertilized treatment (control), the ratios of SOC content in WSA_{mi} to that in soil was 0.80% in the PR + NPK treatment 0.83% and 0.91% in the NPK treatment. The ratios of SOC in WSA, /SOC in bulk soil in the NPK fertilized treatment and in PR + NPK were 14% and 4% higher than in the non-fertilized treatment, respectively. These results demonstrated that all treatments of fertilization had a rather high ability to the sequestration of SOC from bulk soil to WSA_{mi} compared to non-fertilized treatment. The results of Šimanský and Kováčik (2014) also confirmed the fact that water-stable micro-aggregates can be one of the most important storage for retention of SOM due to fertilization of soils. However, as is shown in Table 1, overall, the ratios of SOC in WSA/SOC in bulk soil were higher in macro-aggregates than micro-aggregates. This means that higher C sequestration was in $WSA_{ma'}$ but fertilization did not have any effect on the increase of it in WSA_{ma}. The ratios of C_{L} in WSA/ C_{L} in bulk soil to that in soil, followed the same trends (P > 0.05) as the ratios of SOC in WSA to that in soil with the lowest values in the WSA_{mi} and the highest in WSA_{ma} in all treatments.

Dynamics of SOC and C_{L} are presented in Tables 2–4. The content of SOC in bulk soil had no significant (P > 0.05) increase in all treatments. Results clearly highlighted the effect of time of sampling in spring and autumn as well as plant influence, especially in labile carbon dynamics in fertilized treatments than in non-fertilized treatment. From the point of view of time sampling, the highest contents of SOC and $\mathrm{C}_{\!\scriptscriptstyle L}$ were determined in autumn than spring (Table 2). It can be connected with gradual increase of biomass in bulk soil. The effect of plants on changes of SOC and C, in all treatments is best described by the second polynomial model (based on R²). The highest statistical significant increase of C₁ in bulk soil was identified in autumn, after the cultivation of peas in treatments with ploughed crop residues together with NPK, as well as in only NPK treatment. Miller et al. (2011) presented that green manure legumes such as pea, improved the availability of nutrients in the soil. The resulting increase in residue production also contributes to SOC (Lal, 2008). Intensification of crop rotation is also a key aspect to the observations of increased SOC levels, and may therefore increase organic C input to the soil (Halvorson et al., 2002). The dynamic changes of SOC in WSA were not significant (Table 3), however, in PR + NPK the SOC in WSA_{mi} increased slightly, but the SOC in WSA_{ma} and WSA_{ma} 0.5-3 mm slightly decreased. According to the results of Triberti et al. (2008) continuous additions of organic materials led to a SOC build up at rates 0.16-0.26 t SOC ha-1 year-1. This slow accumulation, which

represented less than 10% of added organic C, can be explained by the great amount of C that annually leaves the soil to other sinks (Richter et al., 2007). Diametrically different situations were in NPK treatment. There were observed slight decreases of SOC in WSA_{mi} and slight increases of SOC in WSA_{ma}. In fertilized treatments, the statistical significant changes in dynamics of C_L in WSA_{ma} and in WSA_{ma} 0.5–3 mm were observed. In PR + NPK there were observed significant decreases because in 2008, the C₁ in WSA_{ma} was 1892 mg kg⁻¹ and in 2011 its content was only 1469 mg kg⁻¹. The same trends were in WSA 0.5-3 mm as well as in the treatment with added NPK fertilizers. A possible reason for the decrease of C_{L} in WSA_{ma} is the newly formed bonds between more labile organic substances and mineral components (Santos et al., 1997) with physical protection against a microbial attack (Krol et al., 2013). There were no significant changes in dynamics of the ratio SOC in WSA/SOC in bulk soil (Table 4). In PR + NPK treatment, the ratio of SOC in WSA_m/SOC in bulk soil increased, however, the ratio of WSA_{ma}/SOC in bulk soil decreased. The opposite trends were in NPK treatment as well as in PR + NPK without statistical significance (Table 4). The ratios of C_{L} in WSA_{mi} and also WSA_{ma}/ C_{L} in bulk soil, decreased due to ploughed plant residues together with NPK fertilizers (statistical significant). Also the ratio C_{L} in WSA_{ma}/ C_{L} in soil statistical significant decreased due to added only NPK fertilizers. All these trends best expressed the polynomial model.

4. Conclusions

The results underscore the importance of fertilization in relation with carbon sequestration, mainly in waterstable macro-aggregates, especially in arable Haplic Luvisol and its meaning is emphasized in this study. Whereas the content of SOM, mainly as a result of the application of crop residues together with NPK fertilizers were slightly increased, there is a potential of increase of C sequestration (especially labile carbon) in a waterstable macro-aggregates, which is definitely one way for the elimination of CO, released from the soil to the atmosphere. Water-stable macro-aggregates are able to retain C, so in this regard it will be necessary to pay further attention to their stability. Application of only NPK fertilizers to soil had negative effect, which means that this alternative, from the view of sustainable farming, is not correct for the future. Significant differences in the dynamics of labile carbon in water-stable aggregates indicated the validity of their use as a sensitive indicator of the quality of the soil environment under different fertilization. This information is very important for farmers, because on this basis, they can optimize soil management practices in arable soils, and avoid environmental degradation due to the application of only mineral fertilizers to the soil.

5. Acknowledgement

Author thanks Ross Mallon (Wellington, New Zealand) for improving the English text.

6. References

BALASHOV, E. and BUCHKINA, N. (2011) Impact of shortand long-term agricultural use of chernozem on its quality indicators. In *Int. Agrophys.*, vol. 25, pp. 1–5.

CARPENTER-BOGGS, L. et al. (2003) Soil microbial properties under permanent grass, conventional tillage, and no-till management in South Dakota. In *Soil Tillage Res.*, vol. 71, pp. 15–23. DOI: http://dx.doi.org/10.1016/S0167-1987(02)00158-7

CHENU, C. and PLANTE, A. (2006) Clay-sized organo-mineral complexes in a cultivation chronosequence: revisiting the concept of the "primary organo-mineral complex". In *European Journal of Soil Science*, vol. 56, pp. 596–607.

DOU, F. et al. (2008) Sensitivity of labile soil organic carbon to tillage in wheat-based cropping systems. In *Soil Sci. Soc. Am. J.*, vol. 72, pp. 1445–1453.

DOI: http://dx.doi.org/10.2136/sssaj2007.0230

DZIADOWIEC, H. and GONET, S.S. (1999) *Methodical guidebook for soil organic matter studies*. Prace Komisji Naukowych Polskiego Towarzystwa Gleboznawczego, N. 120, Komisja chemii gleb, Zespół Materii Organicznej Gleb, N II/16. (in Polish). ELLIOTT, E.T. (1986) Aggregate structure and carbon, nitrogen, and phosphorus in native and cultivated soils. *In Soil Sci. Soc. Am. J.*, vol. 50, pp. 627–633.

EUSTERHUES, K. et al. (2003) Stabilization of soil organic matter by interactions with minerals as revealed by mineral dissolution and oxidative oxidation. In *Organic Geochemistry*, vol. 34, pp. 1591–1600.

GAIDA, A.M. et al. (2013) Changes in soil quality associated with tillage system applied. In *International Agrophysics*, vol. 27, 133–141. DOI: http://dx.doi.org/10.2478/v10247-012-0078-7

HALVORSON, A.D. et al. (2002) Tillage, nitrogen and cropping system effects on soil carbon sequestration. In *Soil Sci. Soc. Am. J.*, vol. 66, pp. 906–912.

DOI: http://dx.doi.org/10.2136/sssaj2002.9060

HAYNES, R.J. (2005) Labile organic matter fractions as central components of the quality of agricultural soils: an overview. In *Adv. Agron.*, vol. 85, pp. 221–268.

IUSS Working Group WRB (2006) World reference base for soil resources. In *World Soil Resources Reports no. 103*. Rome: FAO.

KROL, A. et al. (2013) Effects of organic and conventional management on physical properties of soil aggregates. In *International Agrophysics*, vol. 27, pp. 15–21.

DOI: http://dx.doi.org/10.2478/v10247-012-0063-1

KURAKOV, A.V. and KHARIN, S.A. (2012) The Formation of Water-Stable Coprolite Aggregates in Soddy-Podzolic Soils and the Participation of Fungi in This Process. In *Eurasian Soil Science*, vol. 45, pp. 429–434.

DOI: http://dx.doi.org/10.1134/S1064229312040072

LAL, R. 2008. Carbon sequestration. In *Philos. Trans. R. Soc.*, vol. 363, pp. 815–830.

DOI: http://dx.doi.org/10.1098/rstb.2007.2185

LOGINOW, W. et al. (1987) Fractionation of organic carbon based on susceptibility to oxidation. In *Pol. J. Soil Sci.*, vol. 20, pp. 47–52.

MILLAR, C.E. et al. (1962) *Fundamentals of soil science*. New York : John Wiley and Sons.

MILLER, P.R. et al. (2011) Pea green manure management affects organic winter wheat yield and quality in semiarid Montana. In *Can. J. Plant Sci.*, vol. 91, pp. 497–508.

DOI: http://dx.doi.org/10.4141/cjps10109

NEFF, J.C. et al. (2002) Variable effects of nitrogen additions on the stability and turnover of soil carbon. In *Nature*, vol. 419, pp. 915–917. DOI: http://dx.doi.org/10.1038/nature01136

RICHTER, Jr. D.deB. et al. (2007) Long-term soil experiments: keys to managing Earth's rapidly changing ecosystems. In *Soil Sci. Soc. Am. J.*, vol. 71, pp. 266–279.

DOI: http://dx.doi.org/10.2136/sssaj2006.0181

SANTOS, D. et al. (1997) Uniform separation of concentric surface layers from soil aggregates. In. *Soil Sci. Soc. Am. J.*, vol. 61, pp. 720–724.

DOI: http://dx.doi.org/10.2136/sssaj1997.03615995006100030003x ŠIMANSKÝ, V. (2013) Soil organic matter in water-stable aggregates under different soil management practices in a productive vineyard. In *Arch. Agron. Soil Sci.*, vol. 59, pp. 1207– 1214. DOI: http://dx.doi.org/10.1080/03650340.2012.708103

ŠIMANSKÝ, V. et al. (2013) The effect of organic matter on aggregation under different soil management practices in a vineyard in an extremely humid year. In *Catena*, vol. 101, pp. 108–113. DOI: http://dx.doi.org/10.1016/j.catena.2012.10.011

ŠIMANSKÝ, V. and KOVÁČIK, P. (2014) Carbon sequestration and its dynamic in water-stable aggregates. In *Agriculture*, vol. 60, pp. 1–9. DOI: http://dx.doi.org/10.2478/agri-2014-0001

SIX, J. et al. (2002) Stabilization mechanisms of soil organic matter: implications for C-saturation of soils. In *Plant Soil*, vol. 241, pp. 155–176. DOI: http://dx.doi.org/10.1023/A:1016125726789

SIX, J. et al. (2000) Soil macroaggregate turnover and microaggregate formation: a mechanism for C sequestration under no-tillage agriculture. In *Soil Biology & Biochemistry*, vol. 32, pp. 2099–2103.

DOI: http://dx.doi.org/10.1016/S0038-0717(00)00179-6

SIX, J. and JASTROW, J.D. (2002) Organic matter turnover. In: Lal, R. (Ed.) *Encyclopedia of Soil Science*. New York : Marcel Dekker, pp. 936–942.

TISDALL, J.M. and OADES, J.M. (1980) The effect of crop rotation on aggregation in a red-brown earth. In *Australian Journal of Soil Research*, vol. 18, pp. 423–433.

TRIBERTI, L. et al. (2008) Can mineral and organic fertilization help sequestrate carbon dioxide in cropland? In *Eur. J. Agron.*, vol. 29, pp. 13–20. DOI: http://dx.doi.org/10.1016/j.eja.2008.01.009

VADJUNINA, A.F. and KORCHAGINA, Z.A. (1986) *Methods of Study of Soil Physical Properties.* Moscow: Agropromizdat (in Russian).

von LÜTZOW, M. et al. (2008) Stabilization mechanisms of organic matter in four temperate soils: Development and application of a conceptual model. In *Journal of Plant Nutrition and Soil Science*, vol. 171, pp. 111–124.

DOI: http://dx.doi.org/10.1002/jpln.200700047

WHALEN, J.K. and CHANG, C. (2002) Macroaggregate characteristics in cultivated soils after 25 annual manure applications. In *Soil Sci. Soc. Am. J.*, vol. 66, pp. 1637–1647. DOI: http://dx.doi.org/10.2136/sssaj2002.1637