Original Paper

Genome sequence variation in two subspecies of western honeybee, *A. m. carnica and A. m. ligustica*

Minja Zorc[1], Janko Bozic[2], Peter Dovc[1*]

[1] University of Ljubljana, Biotechnical Faculty, Department of Animal Science, Ljubljana, Slovenia

[2] University of Ljubljana, Biotechnical Faculty, Department of Biology, Ljubljana, Slovenia

Populations of western honeybee (*Apis mellifera*) show differences in morphology, physiology and behaviour as a result of adaptation to various habitats. *A. m. carnica*, which inhabits the South-East and Central Europe, and *A. m. ligustica*, which is endemic on Apennine peninsula, represent 2 closely related honeybee subspecies living in the neighbouring climatic regions. In the current study, 3,655,618 polymorphisms were identified from the whole genome sequences of 37 individual drone genomes, from *A. m. carnica* (n=27) and *A. m. ligustica* (n=10). The analysis revealed variation in genes involved in biological pathways associated with energy production and conversion, cell cycle and cytokinesis.

Keywords: *A. m. carnica*, *A. m. ligustica*, genomics, honeybee, whole genome sequencing

1   Introduction

The genus *Apis* consists of at least 9 honeybee species which are all except western honeybee (*Apis mellifera)* native to southern and eastern Asia (Engel, 1999). As the Asian species of honeybee live in mild tropical climate, they have no need to gather large food stores because they can meet their needs throughout the year (Ruttner, 1988). The western honeybee, which is kept by most beekeepers, is native to Europe, Africa and part of Asia, west of the Caspian sea (Webster, 2008). However, due to human activities, it has become naturalized through much of the rest of the world. Populations of honeybee show differences in morphology, physiology and behaviour as a result of adaptation to various habitats (Le Conte et al., 2008; De la Rúa et al., 2009). There are more than 26 distinct Western honeybee subspecies which can be assigned to 5 evolutionary branches: A (South and Central Africa), C (Central and Southeast Europe), M (Western and North Europe), O (Near and Middle East) and Y (Ethiopia) (Cridland et al., 2017). The honeybees of group C, *A. m. carnica, A. m. ligustica, A. m. caucasica* and *A. m. macedonica,* are variable in behaviour and colour and are adapted to various climates (Ruttner, 1988).

A. m. carnica, commonly known as the Carniolan bee, which inhabits the South-East and Central Europe, and *A. m. ligustica*, commonly known as the Italian bee, inhabiting Apennine peninsula, represent 2 closely related honeybee subspecies living in neighbouring climatic regions (Ruttner, 1988) with evidence of admixture in the natural hybridization zone (Dall'Olio et al., 2007). *A. m. ligustica* bees are yellowish-brown and exhibit low swarming tendencies, while *A. m. carnica* are darker in colour, less aggressive, resistant to disease and good house cleaners (Caron et al., 2013). Studies on metabolic rate and thermal limits of both subspecies suggest that *A. m. ligustica* populations can tolerate a high temperature exposure better than *A. m. carnica* populations (Kovac et

---

* **Corresponding Author:** Peter Dovc. Department of Animal Science, Biotechnical Faculty, University of Ljubljana, Groblje 3, SI-1230 Domzale, Slovenia. E-mail: peter.dovc@bf.uni-lj.si. ORCID: https://orcid.org/0000-0002-7418-0054

Faculty of Agrobiology and Food Resources

al., 2014). There are also variations in the waggle dance between the 2 subspecies (Boch, 1957; Bozic et al., 2016). Several experiments showed no differences regarding learning and memory retention in both subspecies (Hoefer et al., 1975; Iqbal et al., 2019). Also, there were no differences between queens of both subspecies morphometrically or in spermathecal expression of antioxidant-encoding genes (*Catalase, GLOD4L, SOD1, TXN2, TXNRD1* and *Vitellogenin*). However, there was a trend for a higher level of *GSTD1* gene expression in the spermathecae of virgin Italian queens compared to virgin Carniolan queens (Gonzalez et al., 2018). Furthermore, there were no statistical differences in spermathecal sperm counts or sperm viability between Carniolan and Italian mated queens (Gonzalez et al., 2018). Proteomic analysis of royal jelly from both subspecies revealed differences in the heterogeneity of the major royal jelly proteins, in particular *MRJP3* (Li et al., 2007). The comparison of enzyme activities of Carniolan and Italian honey bees, both, uninhibited and after incubation with synergists, gave no significant differences for studied enzymes, which leads to the conclusion that the 2 populations are biochemically similar and comparable (Todeschini et al., 2017). However, in an earlier study, Rinkevich et al. (2015) tested the variation in insecticide susceptibility among honeybee subspecies and there was a large variation in sensitivity to several insecticides between *A. m. ligustica* and *A. m. carnica*. The largest variation in sensitivity was observed in neonicotinoid bioassays, where mutations or post-transcriptional modifications of nicotinic acetylcholine receptors can affect neonicotinoid sensitivity. The significant variance was noticed also in pyrethroid toxicity, which indicates variation in P450 metabolism or voltage-gated sodium channel associated with pyrethroid resistance (Rinkevich et al., 2015).

Whole genome sequences of 10 individual genomes of *A. m. ligustica* drones and 27 individual genomes of *A. m. carnica* drones were applied in the current study to identify sequence variation in subspecies. The identified polymorphisms at the whole genome level among both subspecies could be the first step for their differentiation at the molecular level.

2 Material and methods

2.1 DNA extraction and sequencing

Eight drones of *A. m. carnica* were collected in Slovenia in the pupae/nymph stage. The genomic DNA was extracted from the head and thorax of the frozen larvae cut into small pieces with a scalpel. The DNA extraction was performed using E.Z.N.A. Tissue DNA Kit (Omega Bio-Tek, U.S.A.) according to manufacturer's recommendations. The concentration of genomic DNA for DNA sequencing was adjusted to 50 ng/μl. Pair-end sequencing was performed on Illumina NovaSeq 6000 platform (Genewizz, Leipzig, Germany).

2.2 Read mapping

Eight *A. m. carnica* drone pupae, collected in Slovenia, were sequenced with NovaSeq 6000. Paired-end Illumina HiSeq 2500 reads of 29 publicly available drones (19 *A. m. carnica*, 10 *A. m. ligustica*) were obtained from European Nucleotide Archive under accession number PRJNA311274 (Wragg et al., 2016). The average depth of sequencing coverage was calculated using Qualimap tool (Okonechnikov et al., 2016) and in 8 sequenced *A. m. carnica* drones ranged from 56.90 to 73.39 with an average of 62.17. In publicly available sequences, coverage was lower and ranged from 8.86 to 12.52 with an average of 10.21 in 19 samples of *A. m. carnica,* while in 10 samples of *A. m. ligustica* it ranged from 12.58 to 24.27 with an average of 18.73. Quality control of raw reads was performed using the tool Trim Galore! (https://github.com/FelixKrueger/TrimGalore). The reads were trimmed for adapter and low-quality bases. Trimmed reads were then mapped to the latest honeybee reference genome assembly (HAv3.1) using BWA-MEM (Li and Durbin, 2009) read aligner.

2.3 Variant calling and annotation

Variant sites across all samples were identified using haplotype-based variant detector FreeBayes (https://github.com/ekg/freebayes) with the settings suitable for haploid genomes. Multi-sample vcf file was imported to SVS Golden Helix and multi-nucleotide polymorphisms were split into the single nucleotide polymorphism (SNP) representation. There were 4,903,124 positions which were found polymorphic among individuals. Markers with a call rate <0.90 or with minimum allele frequency <0.05 were removed. For the further analysis, 3,655,618 biallelic markers were retained. SNPs were annotated and filtered using SVS Golden Helix, SNP and Variation suite v8.9.0. Variants that were fixed for alternate alleles in *A. m. carnica* versus *A. m. ligustica* (allele frequency$_{pop1}$ <0.15 and allele frequency$_{pop2}$ >0.85) were identified.

Faculty of Agrobiology and Food Resources

2.4 Gene ontology and biological pathways analysis

Bioinformatics tools g:Profiler (Raudvere et al., 2019) and EnrichmentMap (Isserlin et al., 2014) plugin for Cytoscape (Shannon et al., 2003) were used for functional annotation and enrichment analysis of a filtered set of genes. Functional annotation and enrichment analysis were based on the latest honeybee reference genome assembly (HAv3.1). All known honeybee genes were used as a background gene set in the gene enrichment analysis.

3 Results and discussion

The search for polymorphisms in 37 whole genome sequences of drones belonging to 2 honeybee subpopulations, *A. m. carnica* and *A. m. ligustica,* was performed. We identified 16,239,003 polymorphisms, however, after filtering and quality control 3,655,618 remained for further analysis. On average there was 1 SNP per 62 bp with the gaps that ranged from 1 to 348,017 bp in the reference genome. From 3,655,618 biallelic markers, 805,819 were located in intergenic regions (<5kbp upstream and downstream of the first or last exon, respectively), 2,604,295 markers were located in introns, 100,962 in untranslated regions, and 144,542 in exons, of which 18,991 represented missense variants. On average *A. m. ligustica* genotypes differed from the reference genome on 30.9% of polymorphic sites, whereas *A. m. carnica* differed on 32.4% of sites. There were 989 variants that were fixed for alternate alleles in *A. m. carnica* versus *A. m. ligustica.* Of those, 98 missense SNPs within 64 genes were identified. Among SNPs fixed for different alleles in the 2 subpopulations, there were 9.90% missense SNPs while in the whole set of SNPs missense variants presented only 0.52% variants (P-value <0.00001), similarly as already reported by Wallberg et al. (2014). This could be the evidence for natural selection diversifying both subspecies. In 47 out of 64 genes with missense SNPs there was one missense SNP per gene, and in 13 genes there were 2 missense SNPs. However, in 4 genes we identified 3 or more missense SNPs (Table 1).

Table 1 Genes with 3 or more missense SNPs

Gene	Gene description	Functional descriptions	SNP position	Substitution
LOC100576610	mitochondrial ribonuclease P catalytic subunit	tRNA processing	LG9:11992595	p.Val419Ile
			LG9:11992751	p.Val391Ile
			LG9:11993061	p.Glu329Lys
			LG9:11993480	p.His216Arg
			LG9:11993736	p.Ser131Gly
			LG9:11993772	p.Asp119Asn
			LG9:11993931	p.Asn66Asp
LOC102654405	uncharacterized/ Insect allergen related repeat, nitrile-specifier detoxification	insect allergen-related	LG1:13288906	p.Ile12Val
			LG1:13288907	p.Ile12Thr
			LG1:13288936	p.Ile22Val
			LG1:13289205	p.Asn111Lys
			LG1:13289528	p.Ala219Val
LOC100577452	protein brambleberry	immunoglobulin-like domain	LG8:2410535	p.Leu639Val
			LG8:2411156	p.Leu478Ser
			LG8:2411927	p.Gln276Arg
			LG8:2412152	p.His225Tyr
LOC410743	glucose dehydrogenase [FAD, quinone]	oxidation-reduction process	LG1:6858762	p.Ile444Leu
			LG1:6859449	p.Glu215Lys
			LG1:6859907	p.Leu62Pro

The gene with 7 missense SNPs, *LOC100576610,* is mitochondrial ribonuclease P catalytic subunit (mtRNase P; Figure 1). The evolutionary rate for this gene is high (2.08) in the superfamily Apoidea and lower (1.69) at the level of the class Insecta according to OrthoDB v10.1 (Kriventseva et al., 2019).
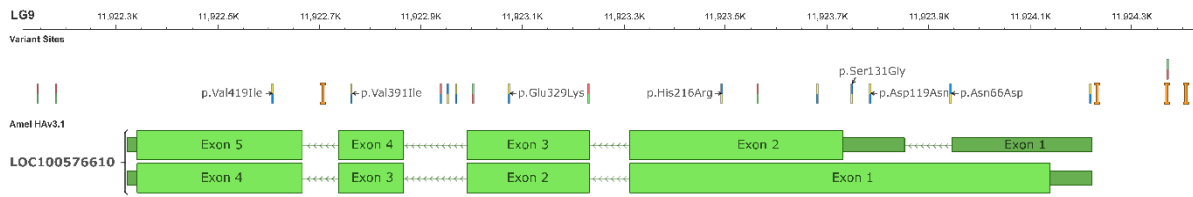
Faculty of Agrobiology and Food Resources

Figure 1 *LOC100576610,* mtRNase P catalytic subunit with 7 missense SNPs

The mtRNase P has been identified as an important factor in RNA processing in a wide number of eukaryotic taxa including yeasts, plants and animals (Gissi et al., 2008). Mitochondrial DNA of the majority of metazoan species encodes only a tiny proportion of proteins (<5%) required for a normal mitochondrial function. Proteins encoded by mitochondrial DNA are components of the enzyme complexes which are responsible for oxidative phosphorylation. Mitochondrial encoded proteins are synthesized by mitochondrial translation machinery, where tRNA plays a crucial role. For a normal tRNA function, cleavage of polycistronic transcripts and specific posttranscriptional modifications of the primary transcripts at both ends are required (Jackman and Alfonzo, 2013). Recently, a specific RNase P (PRORP) has been identified in human mitochondria, being responsible for the 5'-end cleavage of pre-tRNA (Holzmann et al., 2008). In addition to PRORP, the mtRNase P complex contains two additional riboprotein complexes, RNase P Protein 1 and RNase Protein2, both required for selective methylation of tRNA (Vilardo et al., 2012). The PRORP is characterized by the short signal peptide, required for the import of the protein to the mitochondria, repetitive region responsible for RNA binding and the active domain, which is metallonuclease and is responsible for RNA cleaving. The analogue to the human PRORP protein is Mulder protein in *Drosophila melanogaster* (Sen et al., 2016). We compared the coding region for RNase P of *A. m. carnica* and *A. m. ligustica* and identified 7 missense mutations in the coding region suggesting that RNase P is a good candidate locus for molecular discrimination of both subspecies.

Functional enrichment analysis of 64 genes with at least one missense variant revealed enrichment of GO terms (Figure 2) and KEGG pathways associated with energy production and conversion. Enriched KEGG pathways are pyruvate metabolism, fatty acid metabolism, fatty acid elongation, fatty acid degradation, valine, leucine and isoleucine degradation, citrate cycle and glycolysis/gluconeogenesis.
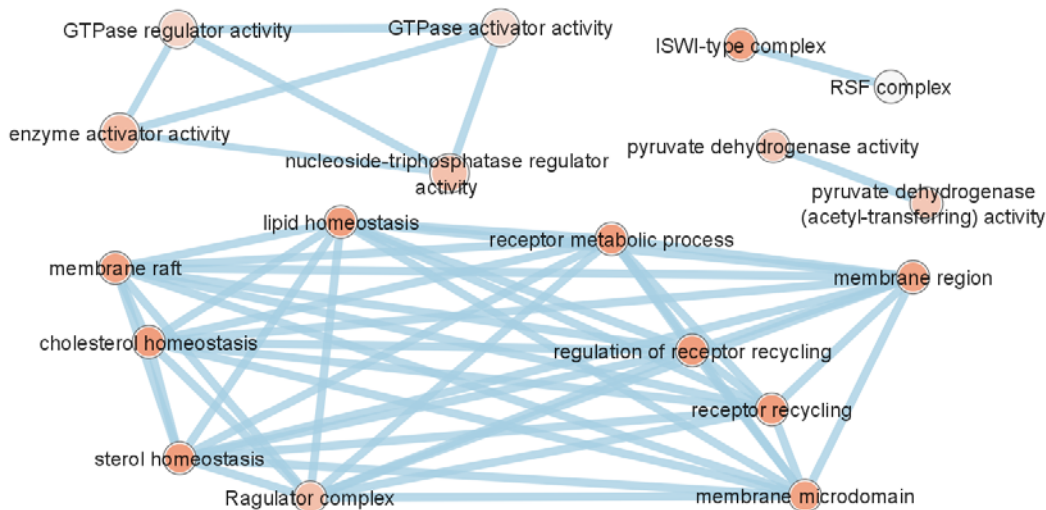

Figure 2 Network representing enriched pathways based on GO terms. Edges represent genes in common between 2 pathways (nodes). The node colour corresponds to the significance of the geneset. Sources GO (BP, MF, CC)

Genes in intersections of enriched biological pathways are ral GTPase-activating protein subunit beta (LOC412130), rab GTPase-binding effector protein 1 (LOC726432), regulator complex protein

LAMTOR1 *(*LOC725789*),* pyruvate dehydrogenase E1 component subunit alpha *(LOC551103)* and trifunctional enzyme subunit beta, mitochondrial *(LOC551775).*

In our material we found 2 missense mutations in the RalGAPbeta subunit between *A. m. carnica* and *A. m. ligustica*, which due to involvement of ral GTPase in different cellular functions may contribute to physiological differences between the 2 subspecies. Members of the RAS-GTPase superfamily were found to have decreased expression in the solid honeybee larval tissues during the development (Chan et al., 2008). The Ral-GTPase is regulated by guanine nucleotide exchange factors and GTPase-activating proteins (GAP) which represent a heterodimeric complex composed of alpha and beta subunit (Shirakawa et al., 2009). RalGAP beta is recruited to the mitotic spindle and intercellular bridges in dividing cells and contributes to the spindle assembly checkpoint. Cells with RalGAPbeta deficiency show altered meta-to anaphase transition in humans (Personnic et al., 2014).

The pyruvate dehydrogenase (PDH) complex is a combination of 3 catalytic components, E1 and E2, which generate acetyl-coenzyme A and FAD/NAD+ dependent E3, which performs redox recycling (Perham, 1991). The PDH catalyses the oxidative decarboxylation of pyruvate and produces acetly CoA, CO2 and NADH(H$^+$). The PDH complex occupies a key position in the oxidation of glucose by linking gycolysis to the cycle of the tricarboxylic acid and maintains the glucose homeostasis (Patel et al., 2014). The PDH catalytic component E1 catalyses 2 consecutive steps, the decarboxylation of pyruvate and the reductive acetylation of the lipoyl groups. The gene encoding PDH belongs to the 3 duplicated glycolysis/gluconeogenesis genes in the honeybee genome (Kunieda et al., 2006).

In our material we found 4 point mutations in the catalytic component E1 of the honeybee PDH. The missense mutations could have an effect on the efficiency of the oxidative decarboxylation of pyruvate also in honeybee and so affect the efficiency of the energy metabolism. Since the activity of the E1 component affects the activity of whole PDH complex, the different genetic variants of the E1 component may play a role in adaptive traits related to energy metabolism. In some cases the deficiency of E1 subunit can cause the demyelination, resulting in neurological symptoms (Singhi et al., 2013).

4   Conclusions

Analysis of genomic sequences of *A. m. carnica* and *A. m. ligustica* revealed differences in the genetic background between the 2 subspecies which may be associated with phenotypic differences in metabolic rate and significant differences in insecticide sensitivity. Analysis of genetic variations in the protein coding part of the genome revealed polymorphisms fixed for different alleles in sampled populations. Among those loci, there was high proportion of missense SNPs, which might be an evidence for natural selection. Functional analysis of genes containing missense SNPs revealed pathways associated with energy metabolism, cell cycle regulation and cytokinesis.

Acknowledgments

References

Boch, R. (1957). Rassenmässige Unterschiede bei den Tänzen der Honigbiene (Apis mellifica L.). *Zeitschrift für vergleichende Physiologie, 40*(3), 289-320. https://doi.org/10.1007/BF00340572

Bozic, J. et al. (2016). Novel aspects in characterisation of Carniolan honey bee *(Apis mellifera carnica*, Pollmann 1879). *Acta Agriculturae Slovenica*, Supplement 5, 18-27.

Caron, D. M. and Connor, L. J. (2013). *Honey Bee Biology and Beekeeping*: Wicas Press.

Chan, Q. W. T. and Foster, L. J. (2008). Changes in protein expression during honey bee larval development. *Genome Biology,* 9(10), R156. https://doi.org/10.1186/gb-2008-9-10-r156

Cridland, J. M., Tsutsui, N. D. and Ramírez, S. R. (2017). The complex demographic history and evolutionary origin of the Western honey bee, Apis Mellifera. *Genome Biology and Evolution*, 9(2), 457-472. https://doi.org/10.1093/gbe/evx009

Dall'Olio, R. et al. (2007). Genetic characterization of Italian honeybees, *Apis mellifera ligustica*, based on microsatellite DNA polymorphisms. *Apidologie,* 38(2), 207-217. https://doi.org/10.1051/apido:2006073

Faculty of Agrobiology and Food Resources

De la Rúa, P. et al. (2009). Biodiversity, conservation and current threats to European honeybees. *Apidologie,* 40(3), 263-284. https://doi.org/10.1051/apido/2009027

Engel, M. S. (1999). The taxonomy of recent and fossil honey bees (Hymenoptera: Apidae; Apis). *Journal of Hymenoptera Research,* 8, 165-196.

Gissi, C., Iannelli, F. and Pesole, G. (2008). Evolution of the mitochondrial genome of Metazoa as exemplified by comparison of congeneric species. *Heredity,* 101(4), 301-320. https://doi.org/10.1038/hdy.2008.62

Gonzalez, A. N., Ing, N. and Rangel, J. (2018). Upregulation of antioxidant genes in the spermathecae of honey bee (*Apis mellifera*) queens after mating. *Apidologie,* 49(2), 224-234. https://doi.org/10.1007/s13592-017-0546-y

Hoefer, I. and Lindauer, M. (1975). Das Lernverhalten zweier Bienenrassen unter veränderten Orientierungsbedingungen. *Journal of Comparative Physiology,* 99(2), 119-138. https://doi.org/10.1007/BF00618179

Holzmann, J. et al. (2008). RNase P without RNA: identification and functional reconstitution of the human mitochondrial tRNA processing enzyme. *Cell,* 135(3), 462-474. https://doi.org/10.1016/j.cell.2008.09.013

Iqbal, J. et al. (2019). Olfactory associative behavioral differences in three honey bee *Apis mellifera L.* races under the arid zone ecosystem of central Saudi Arabia. *Saudi Journal of Biological Sciences,* 26(3), 563-568. https://doi.org/10.1016/j.sjbs.2018.08.002

Isserlin, R. et al. (2014). Enrichment Map - a Cytoscape app to visualize and explore OMICs pathway enrichment results. *F1000Research,* 3, 141. https://doi.org/10.12688/f1000research.4536.1

Jackman, J. E. and Alfonzo, J. D. (2013). Transfer RNA modifications: nature's combinatorial chemistry playground. *Wiley Interdisciplinary Reviews RNA,* 4(1), 35-48. https://doi.org/10.1002/wrna.1144

Kovac, H. et al. (2014). Metabolism and upper thermal limits of *Apis mellifera carnica* and *A. m. ligustica.* *Apidologie,* 45(6), 664-677. https://doi.org/10.1007/s13592-014-0284-3

Kriventseva, E. V. et al. (2019). OrthoDB v10: sampling the diversity of animal, plant, fungal, protist, bacterial and viral genomes for evolutionary and functional annotations of orthologs. *Nucleic Acids Research,* 47(D1), D807-D811. https://doi.org/10.1093/nar/gky1053

Kunieda, T. et al. (2006). Carbohydrate metabolism genes and pathways in insects: insights from the honey bee genome. *Insect Molecular Biology,* 15(5), 563-576. https://doi.org/10.1111/j.1365-2583.2006.00677.x

Le Conte, Y. and Navajas, M. (2008). Climate change: impact on honey bee populations and diseases. *Revue Sentifique et technique,* 27(2), 485-497, 499-510.

Li, H. and Durbin, R. (2009). Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics,* 25(14), 1754-1760. https://doi.org/10.1093/bioinformatics/btp324

Li, J. et al. (2007). Proteomic analysis of royal jelly from three strains of Western Honeybees (*Apis mellifera*). *Journal of Agricultural and Food Chemistry,* 55(21), 8411-8422. https://doi.org/10.1021/jf0717440

Okonechnikov, K., Conesa, A. and Garcia-Alcalde, F. (2016). Qualimap 2: advanced multi-sample quality control for high-throughput sequencing data. *Bioinformatics,* 32(2), 292-294. https://doi.org/10.1093/bioinformatics/btv566

Patel, M. S. et al. (2014). The pyruvate dehydrogenase complexes: structure-based function and regulation. *The Journal of Biological Chemistry,* 289(24), 16615-16623. https://doi.org/10.1074/jbc.R114.563148

Perham, R. N. (1991). Domains, motifs, and linkers in 2-oxo acid dehydrogenase multienzyme complexes: a paradigm in the design of a multifunctional protein. *Biochemistry,* 30(35), 8501-8512. https://doi.org/10.1021/bi00099a001

Personnic, N. et al. (2014). A role for Ral GTPase-activating protein subunit β in mitotic regulation. *The FEBS Journal,* 281(13), 2977-2989. https://doi.org/10.1111/febs.12836

Raudvere, U. et al. (2019). g:Profiler: a web server for functional enrichment analysis and conversions of gene lists (2019 update). *Nucleic Acids Research,* 47(W1), W191-W198. https://doi.org/10.1093/nar/gkz369

Rinkevich, F. D. et al. (2015). Genetics, synergists, and age affect insecticide sensitivity of the honey bee, *Apis mellifera. PloS One,* 10(10), e0139841. https://doi.org/10.1371/journal.pone.0139841

Ruttner, F. (1988). Biogeography and taxonomy of honeybees. Springer-Verlag, Berlin.

Sen, A. et al. (2016). Loss of the mitochondrial protein-only ribonuclease P complex causes aberrant tRNA processing and lethality in Drosophila. *Nucleic Acids Research,* 44(13), 6409-6422. https://doi.org/10.1093/nar/gkw338

Shannon, P. et al. (2003). Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Research,* 13(11), 2498-2504. https://doi.org/10.1101/gr.1239303

Shirakawa, R. et al. (2009). Tuberous sclerosis tumor suppressor complex-like complexes act as GTPase-activating proteins for Ral GTPases. *Journal of Biological Chemistry,* 284(32), 21580-21588. https://doi.org/10.1074/jbc.M109.012112

Singhi, P. et al. (2013). Pyruvate dehydrogenase-e1α deficiency presenting as recurrent demyelination: an unusual presentation and a novel mutation. *JIMD Reports,* 10, 107-111. https://doi.org/10.1007/8904_2012_211

Todeschini, V. et al. (2017). In vitro study on the inhibition of enzymatic systems in Italian and Carniolan honey bees by piperonyl butoxide new derivatives. *Bulletin of Insectology,* 70, 237-244.

Vilardo, E. et al. (2012). A subcomplex of human mitochondrial RNase P is a bifunctional methyltransferase--extensive moonlighting in mitochondrial tRNA biogenesis. *Nucleic Acids Research,* 40(22), 11583-11593. https://doi.org/10.1093/nar/gks910

Wallberg, A. et al (2014). A worldwide survey of genome sequence variation provides insight into the evolutionary history of the honeybee *Apis mellifera. Nature Genetics,* 46(10), 1081-1088. https://doi.org/10.1038/ng.3077

Webster, T. C. (2008). Honey Bee, *Apis mellifera* (Hymenoptera: Apidae). In J. L. Capinera (Ed.), *Encyclopedia of Entomology* (pp. 1835-1840). Dordrecht: Springer Netherlands.

Wragg, D. et al. (2016). Whole-genome resequencing of honeybee drones to detect genomic selection in a population managed for royal jelly. *Scientific Reports,* 6, 27168. https://doi.org/10.1038/srep27168

Faculty of Agrobiology and Food Resources